THE PHOTOOXYGENATION OF BILIVERDIN

David A. Lightner and Dave C. Crandall<sup>†</sup> Department of Chemistry, Texas Tech University Lubbock, Texas 79409

(Received in USA 30 January 1973; received in UK for publication 10 February 1973)

The extensively used phototherapy for neonatal jaundice has been widely discussed<sup>1,2</sup>, and its success presumably depends at least in part on the photodegradation of bilirubin IXa (]) to excretable, water-soluble products. McDonagh has shown that bilirubin is a singlet oxygen  $({}^{1}0_{2})$  sensitizer<sup>3</sup>, and Bonnett and Stewart have recently reinforced this belief<sup>4</sup>. Some of the water soluble mono- and dipyrroles from <u>in vitro</u> photooxidation of ] have been identified<sup>5,6,7</sup>. Ostrow<sup>8,9,10</sup> and Schmid<sup>10</sup> have also indicated that the <u>in vitro</u> photo-destruction of ] gives biliverdin (2a), and Ostrow<sup>9</sup> has postulated that 2a is an intermediate in the photooxidation of ]. In our work on the photooxidation of ] we have identified 2a as a photoproduct<sup>11</sup>, but we do not believe that 2a is the precursor to photoproducts of ], for 2a is photodestroyed at a considerably slower rate than is  $1^{12}$ . Furthermore, McDonagh has shown that 2 is a singlet oxygen quencher<sup>13</sup>. Despite these findings, very little is know of the photooxidation products of  $2a^{14}$ . We wish to report here on the first isolation and structure proof of hematinic acid methyl ester (3) and of a methanol propentdyopent adduct (4) from photooxidation of biliverdin dimethyl ester (2b).

Crude biliverdin IXa (2a) was prepared in 90% yield by controlled oxidation of bilirubin IXa (1) [100 mg, Matheson, Coleman and Bell] using benzoquinone (101 mg) in 50 ml of dimethyl-sulfoxide-acetic acid (9:1 v/v) according to the procedure of Bonnett and McDonagh<sup>15</sup>. The pure dimethyl ester (2b) was prepared from 2 (22% overall yield from 1) by esterification using  $BF_3$ -methanol<sup>15</sup> followed by preparative thin layer chromatography (Silica Gel F, M. Woelm,

<sup>\*</sup>For preceding paper see D. A. Lightner and G. B. Quistad, Fed. Europ. Biochem. Soc. (FEBS) Lett., 25, 94 (1972).

<sup>&</sup>lt;sup>T</sup>National Science Foundation Undergraduate Research Participant (1971-72) and President's Undergraduate Fellow at the University of California.

Eschwege, 1mm, 12% acetone in CHC1<sub>3</sub> v/v). Pure 2b moved as a dark blue-green band, R<sub>f</sub> 0.46, with only slight traces of the III<sub>a</sub> and XIII<sub>a</sub> isomers.<sup>16</sup>

A dilute (1.88 mM) methanolic solution of biliverdin IX $\alpha$  dimethyl ester (2b) containing 4.3 mg % Rose Bengal ( $^{1}O_{2}$  sensitizer) was irradiated in an immersion well apparatus using 500 watt Westinghouse tungsten-halogen lamp (500 Q/CL) at 100V while bubbling a slow stream of oxygen through the solution. After a photooxidation period of 32 hours (followed in the ultraviolet-visible spectrum by the disappearance of the 670 and 380 nm maxima of 2b and the emergence of new maxima at 570, 310 and 280 nm), the photoproducts were separated by a combination of gradient elution column chromatography on silica gel (E. Merck, Darmstadt, 70-325 mesh ASTM) using ether ightarrow ethyl acetate ightarrow acetone ightarrow methanol and preparative tlc (Silica Gel F, M. Woelm, Eschwege, 1mm, ether). Two new substances were isolated from a polar mixture. The structure of the methanol propentdyopent adduct methyl ester (3) isolated in 10% yield, was determined by its mass spectrum  $\frac{17}{m/e}$  (relative intensity) 346.1526 (10%) [M<sup>+</sup>, C<sub>10</sub>H<sub>22</sub>N<sub>2</sub>O<sub>F</sub>], 315 (9%) [M-OCH3], 314 (17%) [M-HOCH3], 259 (14%) [M-CH2CO2CH3], 256 (44%) [M-CO2CH3], 256 (44%) 212 (75%) and 152 (100%); its nmr spectrum:  $\delta$  1.80 (CH<sub>3</sub>/s), 1.95 (CH<sub>3</sub>/s), 2.59 (-CH<sub>2</sub>CH<sub>2</sub>-/s), 3.12 (OCH<sub>3</sub>/s), 3.62 (CO<sub>2</sub>CH<sub>3</sub>/s), 4.76 (-CH=, meso/s), 5.59 (-CH=, viny1/m), 6.30 (=CH<sub>2</sub>/m), and 8.30 (NH/br. s) ppm; and its ultraviolet spectrum:  $\epsilon_{293 nm}$  14,000 methanol. A positive pentdyopent reaction,  $\lambda_{max} = 524$  nm, was obtained for 3.18 The expected isomeric compound  $(4)^5$  was not found. We presume it suffers more rapid (than 3) non-photochemical alterations, as we have shown that the hydroxy isomer of 4 is much more labile than the hydroxy isomer of 3 upon standing in methanol at room temperature<sup>19</sup>. Hematinic acid methyl ester (5) was isolated in 5% yield. Its structure was proved by its mass spectrum, m/e (relative intensity): 197.0681 (4%)  $[M^+, C_0H_{11}NO_4]$ , 165 (100%)  $[M-CH_3OH]$ , and 137 (85%)  $[M-CO_2CH_3-H]$ ; and its nmr spectrum: & 1.99 (CH<sub>3</sub>/s), 2.66 (-CH<sub>2</sub>-CH<sub>2</sub>, s), 3.65 (OCH<sub>3</sub>/s), and 7.52 (NH/br. s) ppm.

As shown in Scheme 1, we believe that both 3 and 5 are formed from an endoperoxide intermediate A (Path I) involving the inner rings of 2b with subsequent cleavages reminiscent of those observed for 3,4-diethyl-2,5-dimethylpyrrole<sup>20</sup>. These findings may be coupled with the earlier report of isolating methylvinylmaleimide (6) from unsensitized photooxygenation of 2a, presumably via a dioxetane intermediate (B) formed (Path II) by addition of  ${}^{10}O_{2}$  to the enamine-like bridges a and c as originally suggested by McDonagh<sup>3</sup> based on the earlier observations of Foote and Lin<sup>21</sup> for enamines. Thus, two different modes of attack by  ${}^{10}O_{2}$  on the biliverdin skeleton are clearly implicated (Routes I and II), just as they are for bilirubin<sup>5</sup>. We are currently looking for additional photoproducts from 2a and 2b.

<u>Acknowledgement</u>: The authors gratefully thank the National Institute of Child Health and Human Development, U.S. Public Health Service (HD-07358) and the National Science Foundation (GP-32483X) for support of this work. All mass spectra reported were run by Miss Elizabeth Irwin of the UCLA Chemistry Department. We wish to thank Dr. Gary B. Quistad for helpful and stimulating discussions during the course of this work.





## REFERENCES

- For a recent summary see D. Bergsma, D. Y.-Y. Hsia and C. Jackson, eds., <u>Bilirubin</u> <u>Metabolism of the Newborn</u>, Williams and Wilkins Co., Baltimore, 1970.
- R. Schmid, <u>New Eng. J. Med.</u>, 285, 520 (1971). Also, see leading articles in <u>Brit. Med.</u> <u>J.</u>, 2, 5 (1970); <u>Lancet</u>, 1, 825 (1970); and J. Pediatrics, 74, 989 (1969).
- 3. A. F. McDonagh, <u>Biochem. Biophys. Res. Commun.</u>, <u>44</u>, 1306 (1971).
- 4. R. Bonnett and J. C. M. Stewart, <u>Biochem. J.</u>, <u>J30</u>, 895 (1972).
- 5. D. A. Lightner and G. B. Quistad, Fed. Europ. Biochem. Soc. (FEBS) Lett., 25, 94 (1972).
- 6. R. Bonnett and J. C. M. Stewart, <u>Chem. Commun.</u>, 596 (1972).
- 7. C. H. Gray, A. Kulcyzka and D. C. Nicholson, J. Chem. Soc., Perkin I, 288 (1972).
- J. D. Ostrow in <u>Bilirubin Metabolism</u>, J. A. D. Bouchier and B. H. Billings, eds., Blackwell Scientific, Oxford, 1967, page 117.
- 9. J. D. Ostrow and R. V. Branham, <u>Gastroenterology</u>, 58, 15 (1970) and references therein.
- 10. J. D. Ostrow, L. Hammaker and R. Schmid, <u>J. Clin. Invest.</u>, <u>40</u>, 1442 (1961).
- 11. D. A. Lightner and G. B. Quistad, <u>Nature New Biology</u>, 236, 203 (1972).
- 12. D. A. Lightner, D. C. Crandall, S. Gertler and G. B. Quistad, unpublished data.
- 13. A. F. McDonagh, <u>Biochem. Biophys. Res. Commun.</u>, <u>48</u>, 408 (1972).
- D. A. Lightner and D. C. Crandall, Fed. Europ. Biochem. Soc. (FEBS) Lett., 20, 53 (1972).
- R. Bonnett and A. F. McDonagh, <u>Chem. Commun.</u>, 238 (1970). We thank Dr. A. F. McDonagh for details of this procedure.
- 16. A. F. McDonagh and F. Assisi, Fed. Europ. Biochem. Soc. (FEBS) Lett., 18, 315 (1971).
- 17. Mass spectra were determined on a CEC MS 491-21 or an AEI MS-9 mass spectrometer, nuclear magnetic resonance (nmr) spectra were run on a Varian T-60 instrument using CEC1<sub>3</sub> solvent, and visible-ultraviolet spectra were recorded on a Cary 14 spectrophotometer.
- H. von Dobeneck. <u>Z. Clin. Chem.</u>, 4, 137 (1966); R. Bonnett, M. J. Dimsdale and G. F. Stephenson, <u>Chem. Commun.</u>, 1121 (1968).
- 19. G. B. Quistad and D. A. Lightner, unpublished observation.
- 20. D. A. Lightner and G. B. Quistad, Angew. Chemie, 84, 216 (1972).
- 21. C. S. Foote and J. W.-P. Lin, <u>Tetrahedron Lett</u>., 3267 (1968).